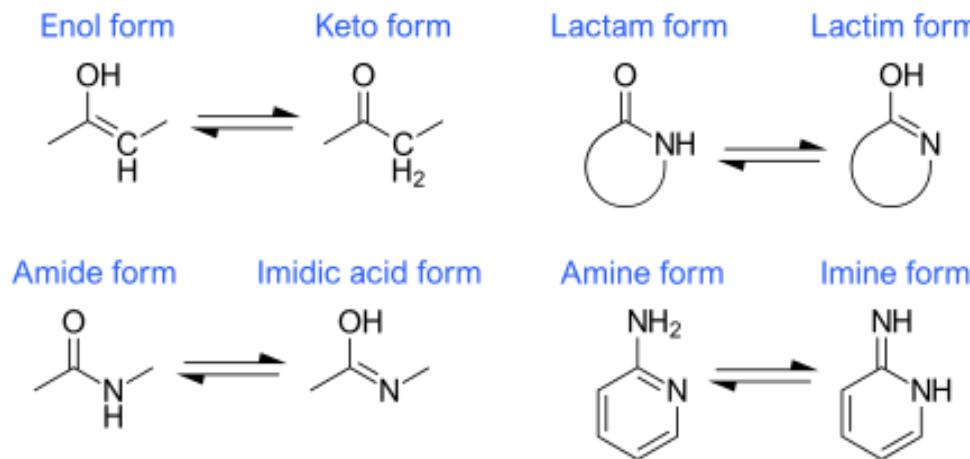


Other phosphorylated compounds and thioesters also have large free energy of hydrolysis

- ▶ **Tautomerization** (호변이성)
; Phosphoenolpyruvate

Tautomers are constitutional isomers of organic compounds that readily interconvert. This reaction commonly results in the relocation of a proton

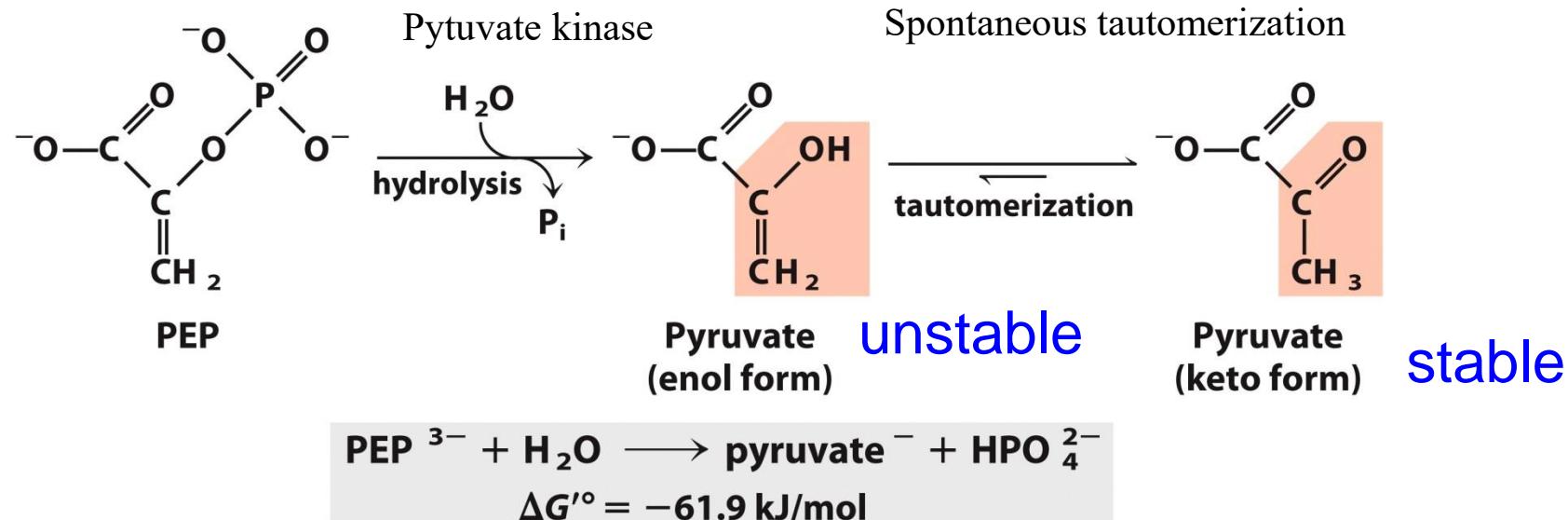


- ▶ **Resonance stabilization** by removing Pi
; 1,3-biphosphoglycerate, phosphocreatine
- ▶ **Thioester**

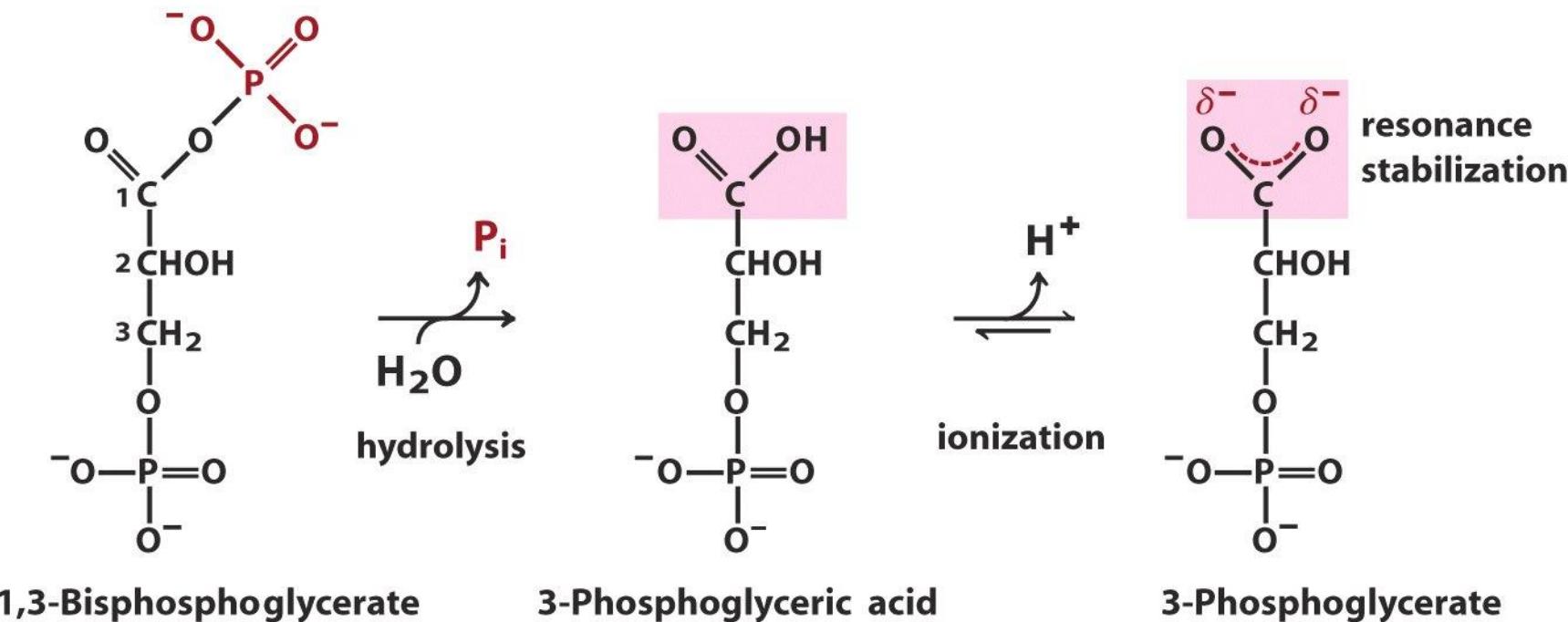
Several phosphorylated compounds have large $\Delta G^\circ'$ for hydrolysis

1. Hydrolysis of phosphoenolpyruvate (PEP)

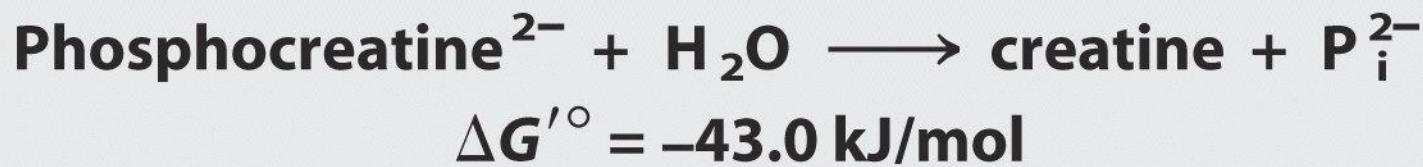
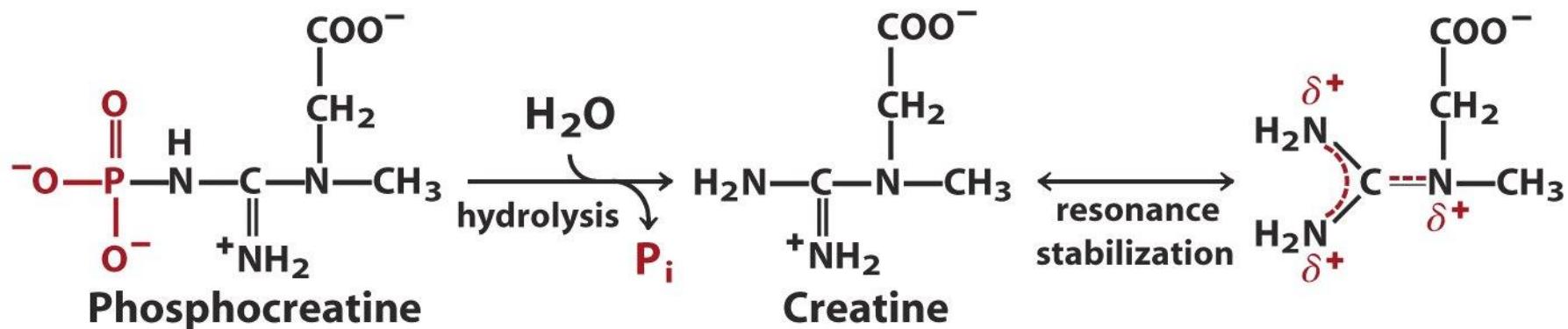
- Electrostatic repulsion within the reactant molecule is relieved
- The products (P_i) are stabilized via resonance, or by more favorable solvation
- The product undergoes further tautomerization



2. Hydrolysis of 1,3-bisphosphoglycerate (1, 3,BPG)



3. Hydrolysis of phosphocreatine



4. Hydrolysis of acetyl-Coenzyme A

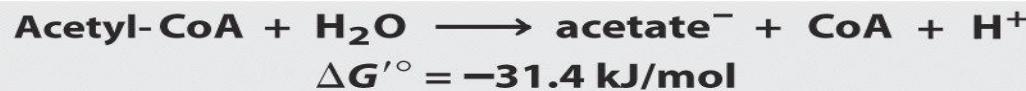
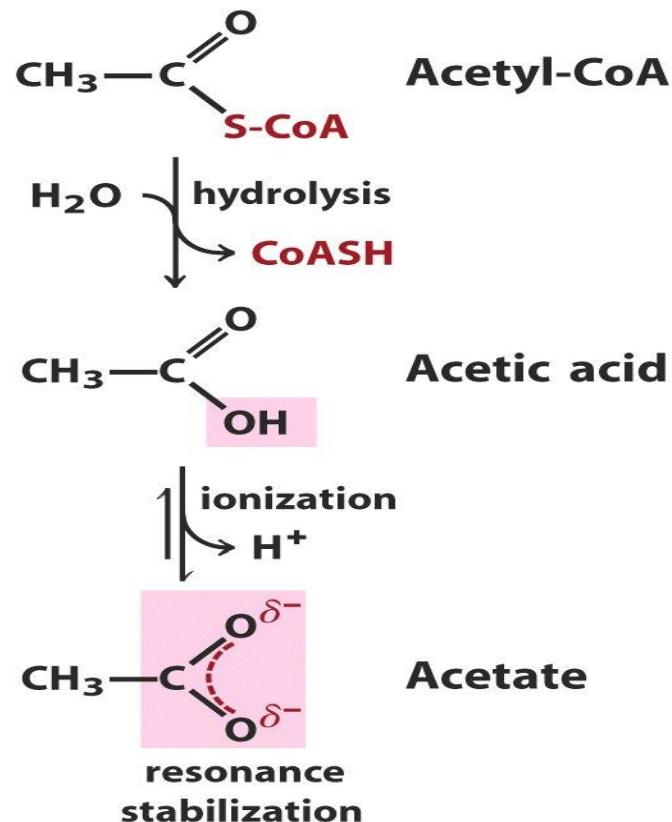


TABLE 13–6 Standard Free Energies of Hydrolysis of Some Phosphorylated Compounds and Acetyl-CoA (a Thioester)

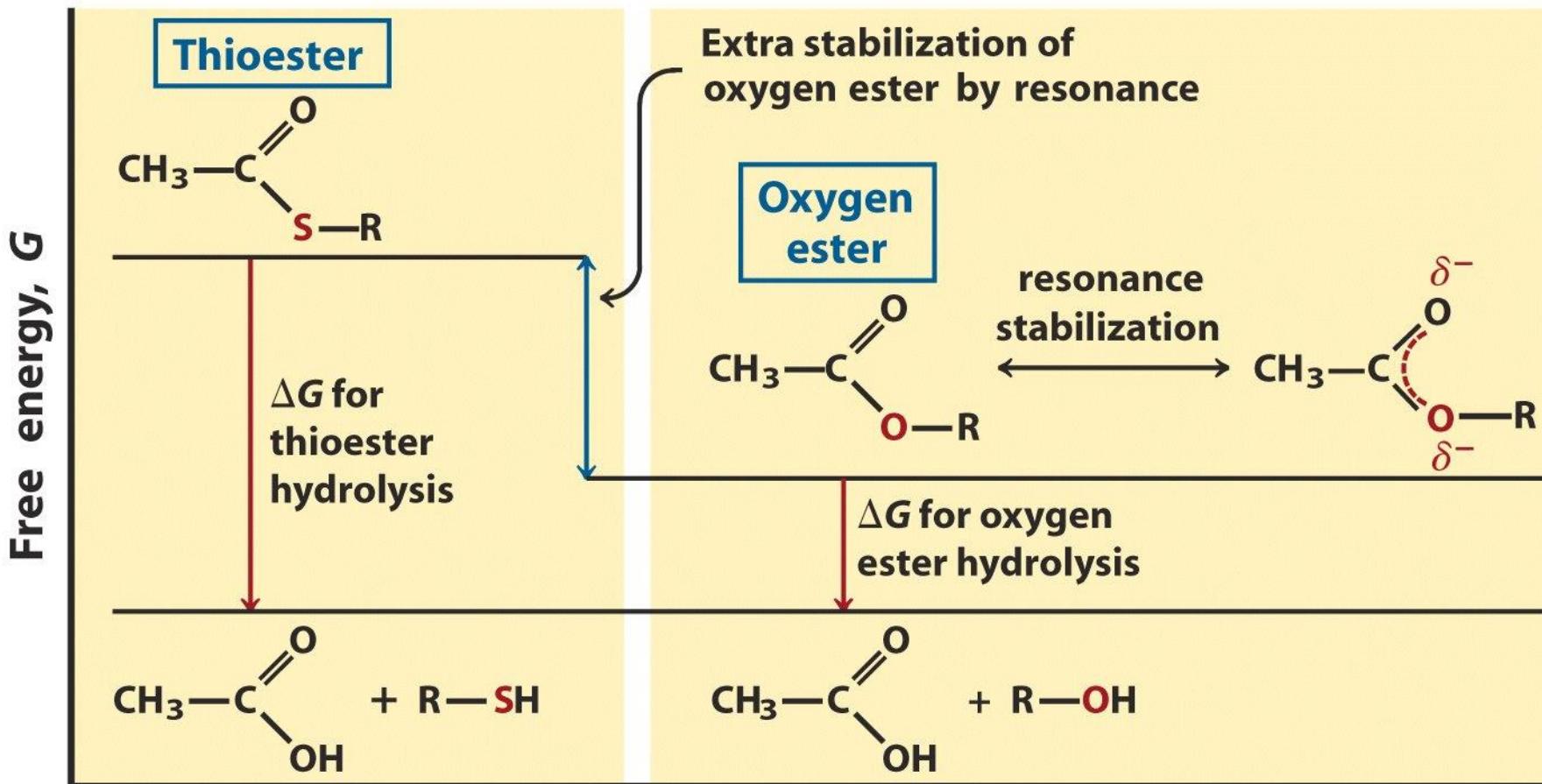
| | $\Delta G'^\circ$ | |
|---|-------------------|------------|
| | (kJ/mol) | (kcal/mol) |
| Phosphoenolpyruvate | −61.9 | −14.8 |
| 1,3-bisphosphoglycerate (\rightarrow 3-phosphoglycerate + P_i) | −49.3 | −11.8 |
| Phosphocreatine | −43.0 | −10.3 |
| ADP (\rightarrow AMP + P_i) | −32.8 | −7.8 |
| ATP (\rightarrow ADP + P_i) | −30.5 | −7.3 |
| ATP (\rightarrow AMP + PP_i) | −45.6 | −10.9 |
| AMP (\rightarrow adenosine + P_i) | −14.2 | −3.4 |
| PP_i (\rightarrow $2P_i$) | −19.2 | −4.0 |
| Glucose 1-phosphate | −20.9 | −5.0 |
| Fructose 6-phosphate | −15.9 | −3.8 |
| Glucose 6-phosphate | −13.8 | −3.3 |
| Glycerol 1-phosphate | −9.2 | −2.2 |
| Acetyl-CoA | −31.4 | −7.5 |

Source: Data mostly from Jencks, W.P. (1976) in *Handbook of Biochemistry and Molecular Biology*, 3rd edn (Fasman, G.D., ed.), *Physical and Chemical Data*, Vol. I, pp. 296–304, CRC Press, Boca Raton, FL. The value for the free energy of hydrolysis of PP_i is from Frey, P.A. & Arabshahi, A. (1995) Standard free-energy change for the hydrolysis of the α - β -phosphoanhydride bridge in ATP. *Biochemistry* **34**, 11,307–11,310.

Free energy of hydrolysis for thioester and oxygen esters

acetyl-CoA; donor of acyl groups, Feeding two-carbon units into metabolic pathways

Synthesis of fatty acids

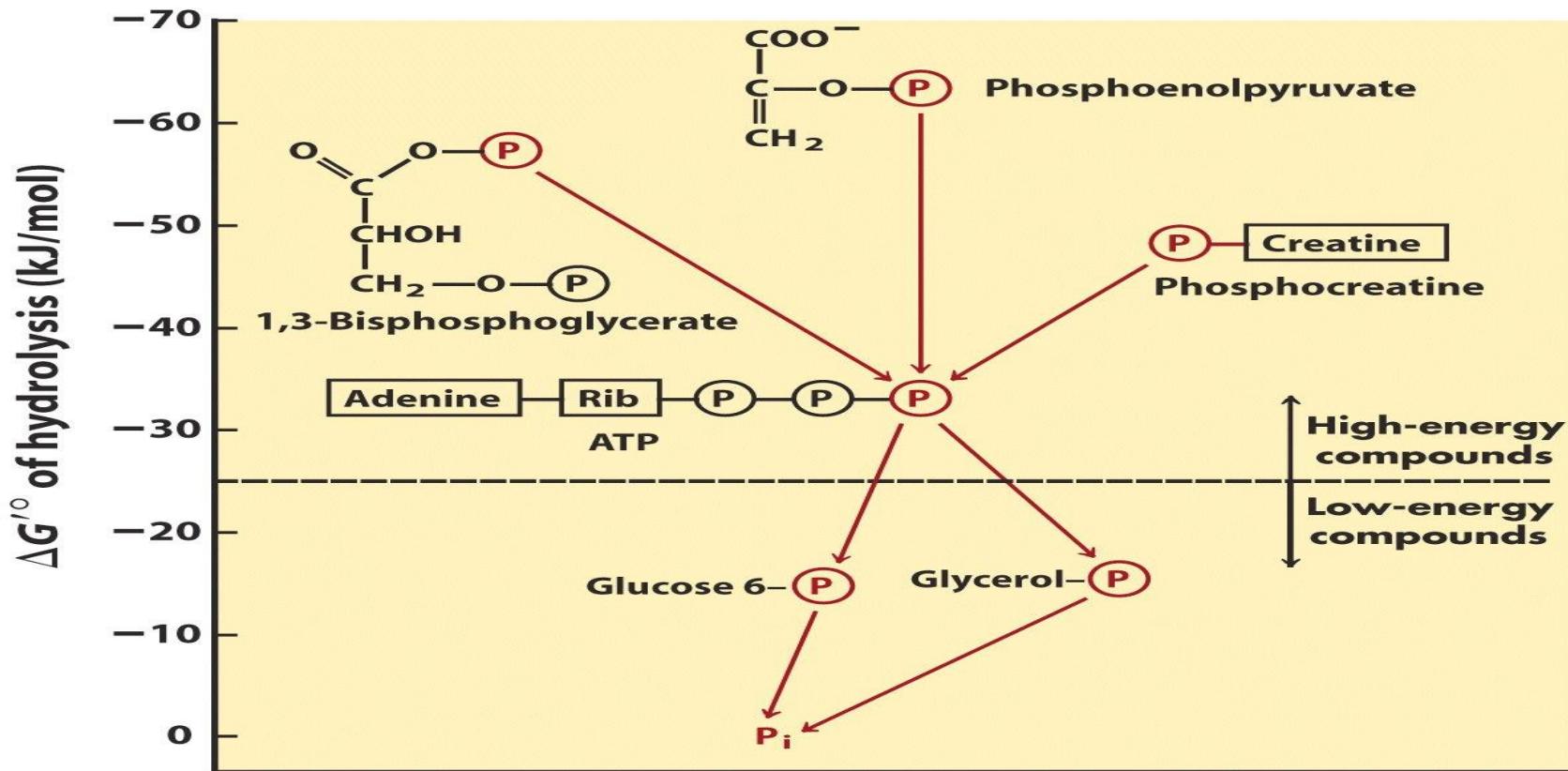


Summary for hydrolysis reactions with large, negative ΔG°

The products are more stable than the reactant by:

- (1) charge separation relieve electrostatic repulsion
- (2) product stabilization by ionization
- (3) by isomerization (tautomerization)
- (4) by resonance stabilization

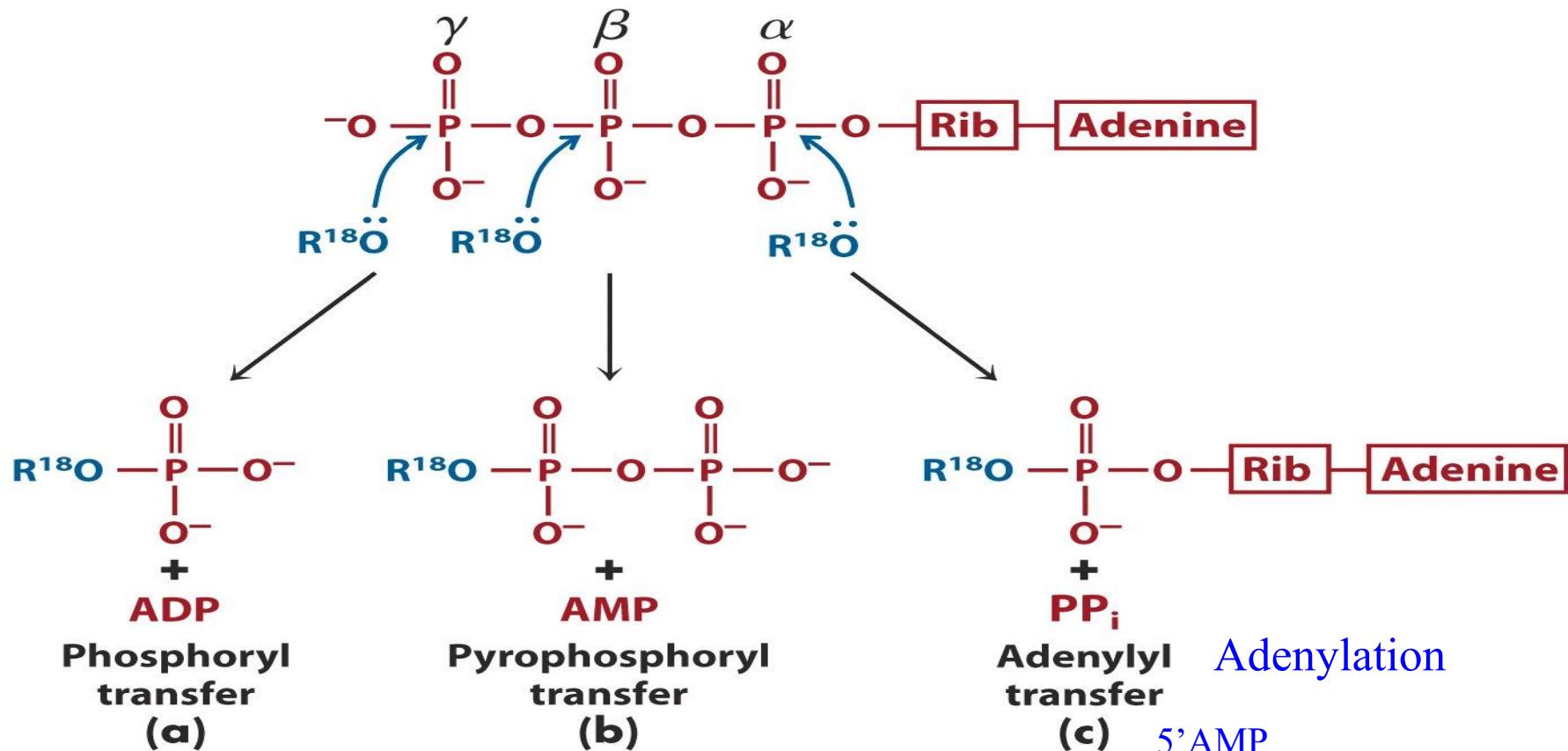
Ranking of biological phosphate compounds by standard free energy of hydrolysis



ATP: Intermediate position on the scale of group transfer potential
(Universal energy currency)

ATP donates phosphoryl, Pyrophosphoryl, and Adenyl group

Three positions on ATP for attack by the nucleophile $R^{18}O$



The nucleophile may be an alcohol ($R-OH$), a carboxy group ($R-COOH$), or a phosphoanhydride

The oxidation levels of carbon in biomolecules

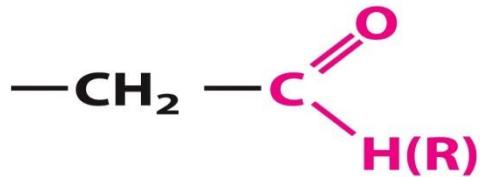
Reduced organic compounds serve as **fuels** from which electrons can be stripped off during oxidation.



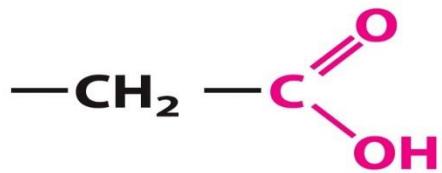
Alkane



Alcohol



Aldehyde (ketone)



Carboxylic acid



Carbon dioxide

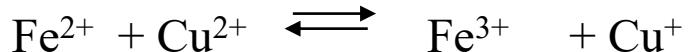
Each compound is formed by oxidation of the red carbon in the compound shown immediately above. **Carbon dioxide is the most highly oxidized form of carbon found in living systems.**

Biological oxidation-reduction reactions

- ▶ Transfer of phosphoryl group
- ▶ Electron transfer in oxidation-reduction reactions
 - Loss of electron: oxidized
 - Gain of electron: reduced
- ▶ Source of electron
 - In non-photosynthetic (Food) vs photosynthetic (excited by light)
- ▶ Electrons **move** from metabolic intermediates to **specialized electron carriers** in enzyme-catalyzed reactions
- ▶ The flow of electron can do biological work
 - Electromotive force (emf) → proton-motive force → ATP (chemical work)

Oxidation-Reduction can be described as half-reaction

- Free energy change is also associated with equilibrium with oxidation-reduction reaction
- Oxidation-reduction reaction occur together. It is convenient when describing e- transfer to consider the two halves of an oxidation-reduction reaction separately.



; In acid-base reaction, proton donor \rightleftharpoons proton acceptor + H +
An acid and corresponding base function as a conjugated acid-base pair.

; In redox reaction, e- donor (reductant) \rightleftharpoons e- acceptor (oxidant) + e-

- e- donor and corresponding e- acceptor function as a conjugated redox pair

(The e- donating molecules in oxidation-reduction reaction is reducing agent or reductant
The e- accepting molecules is oxidizing agent or oxidant)

Reduction potentials measure affinity for electrons

- ▶ When two conjugate redox pairs are together in solution, e- transfer from e- donor of one pair to e- acceptor of other, may proceed spontaneously. The tendency for such a reaction depends on **relative affinity of e-acceptor** of each redox pair **for electrons**.
- ▶ **Redox couple: couple of oxidant and reductant**
- ▶ **Standard reduction potential (표준환원전위, E°)**: a measure (in volts) of this affinity (measure in which e- donor loss electron ; electron donor 가 전자를 잃어버리는 경향의 척도) ; Redox couple with large negative E° provide electron redox couple with large positive E°
- ▶ $\Delta G^\circ = -n F \Delta E^\circ$

n: the number of e- transferred in the reaction

F: faraday's constant

ΔE° : standard reduction potential at pH 7

Standard reduction potentials can be used to calculate free-energy change

► Why are reduction potential so useful to the biochemist?

; We can predict the direction in which electron will tend to flow when components of both half-cells are present in the same solution.

► Electron tend to flow to half-cell with more positive E and strength of that tendency is proportional to ΔE , difference in reduction potential.

► The energy made available by spontaneous e- flow (free energy change, ΔG) is proportional to ΔE

- $\Delta G^\circ = -n F \Delta E^\circ$

n: the number of e- transferred in the reaction

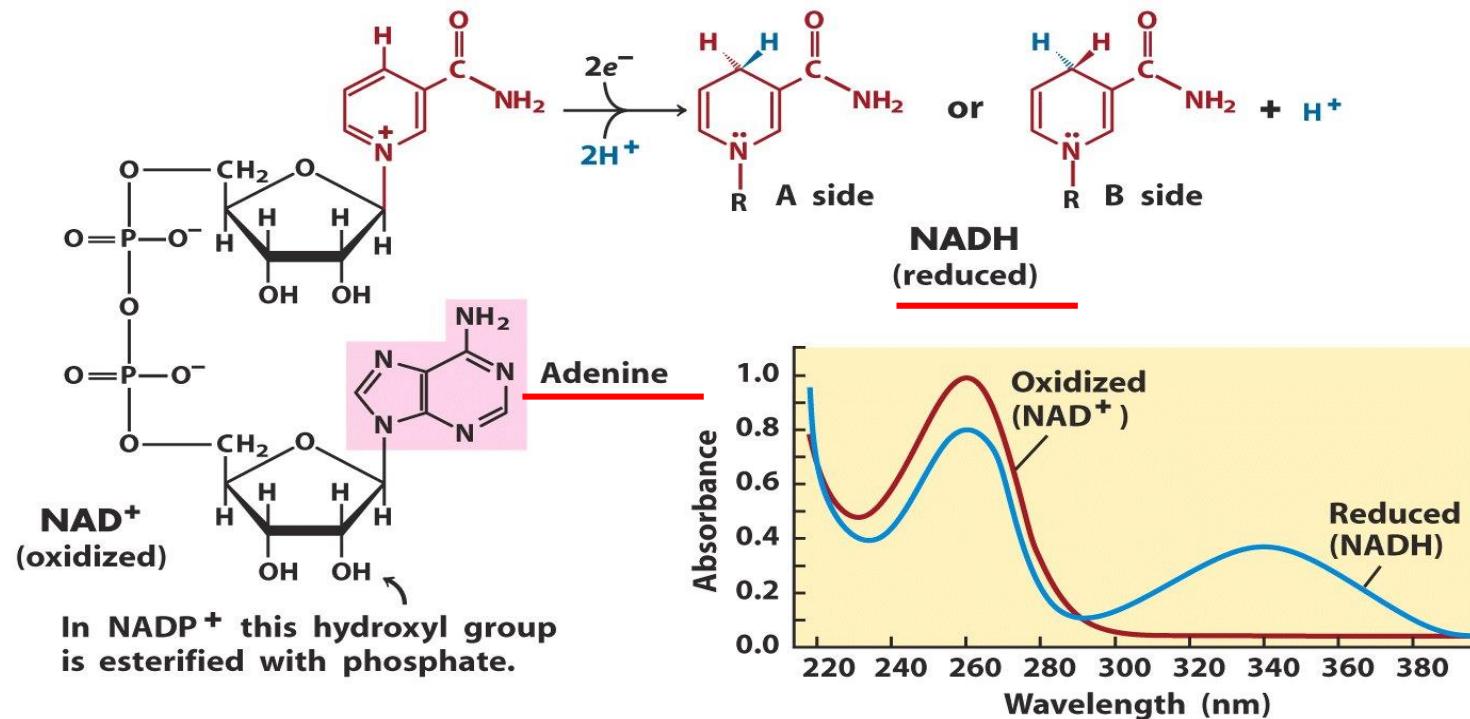
F: faraday's constant

ΔE° : standard reduction potential at pH 7

► Redox couple with large negative E° provide electron redox couple with large positive E°

- Cellular oxidation of glucose to carbon dioxide requires specialized electron carriers
- A few types of coenzymes and proteins serve as universal electron carriers
- **NADH** and **NADPH** act with dehydrogenase as soluble electron carriers

Hydride ion is added to either front side (A side) or back side (B side)



Nicotinamide adenine dinucleotide (phosphate) (NAD (P))

In catabolism, reducing equivalent is released from these substrate, often in the form of hydride ion (a proton coupled with two electron, H:-)

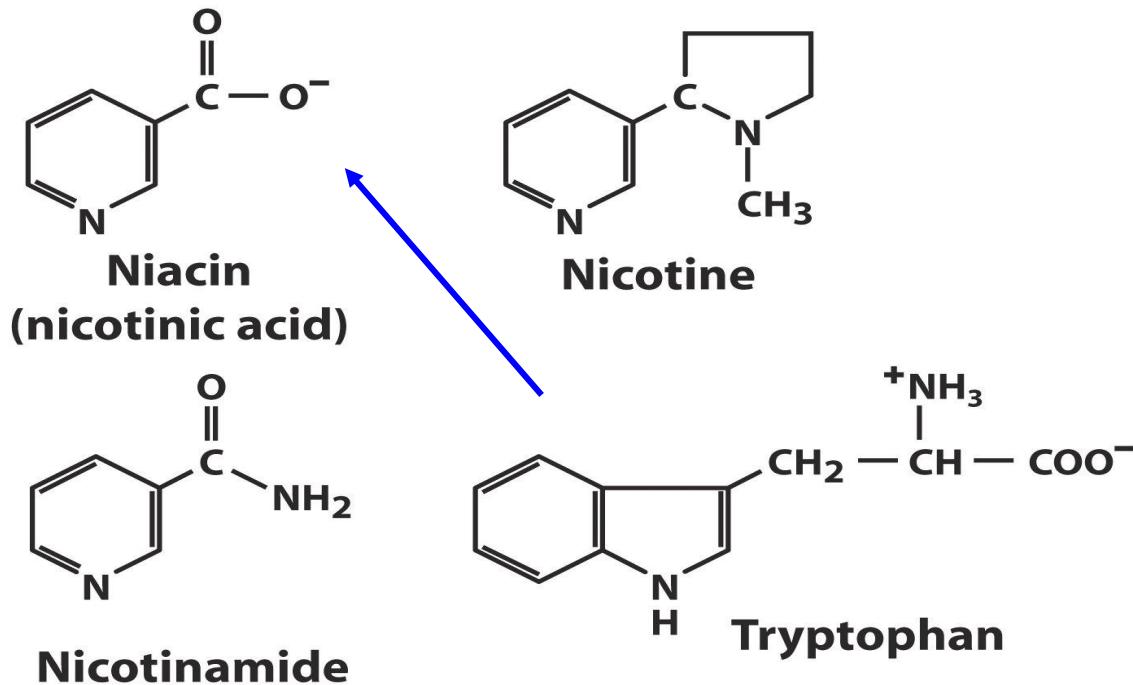
TABLE 13-8 Stereospecificity of Dehydrogenases That Employ NAD⁺ or NADP⁺ as Coenzymes

| Enzyme | Coenzyme | Stereochemical specificity for nicotinamide ring (A or B) | Text page(s) |
|--|---------------------------------------|---|--------------|
| Isocitrate dehydrogenase | NAD ⁺ | A | 610 |
| α -Ketoglutarate dehydrogenase | NAD ⁺ | B | 610 |
| Glucose 6-phosphate dehydrogenase | NADP ⁺ | B | 540 |
| Malate dehydrogenase | NAD ⁺ | A | 612 |
| Glutamate dehydrogenase | NAD ⁺ or NADP ⁺ | B | 665 |
| Glyceraldehyde 3-phosphate dehydrogenase | NAD ⁺ | B | 530 |
| Lactate dehydrogenase | NAD ⁺ | A | 538 |
| Alcohol dehydrogenase | NAD ⁺ | A | 540 |

Dietary deficiency of niacin, the vitamin form of NAD and NADP, cause pellagra

- Pellagra (rough skin): dermatitis, diarrhea, dementia
→ followed in many case by death
- Individual with **diets low in tryptophan content (maize)** has **niacin deficiency**,

which affect all the NAD(P)-dependent dehydrogenase, causes the serious human disease Pellagra (Italian for **rough skin**)



Flavin nucleotide are tightly bound in flavoproteins

Oxidized and reduced FAD and FMN

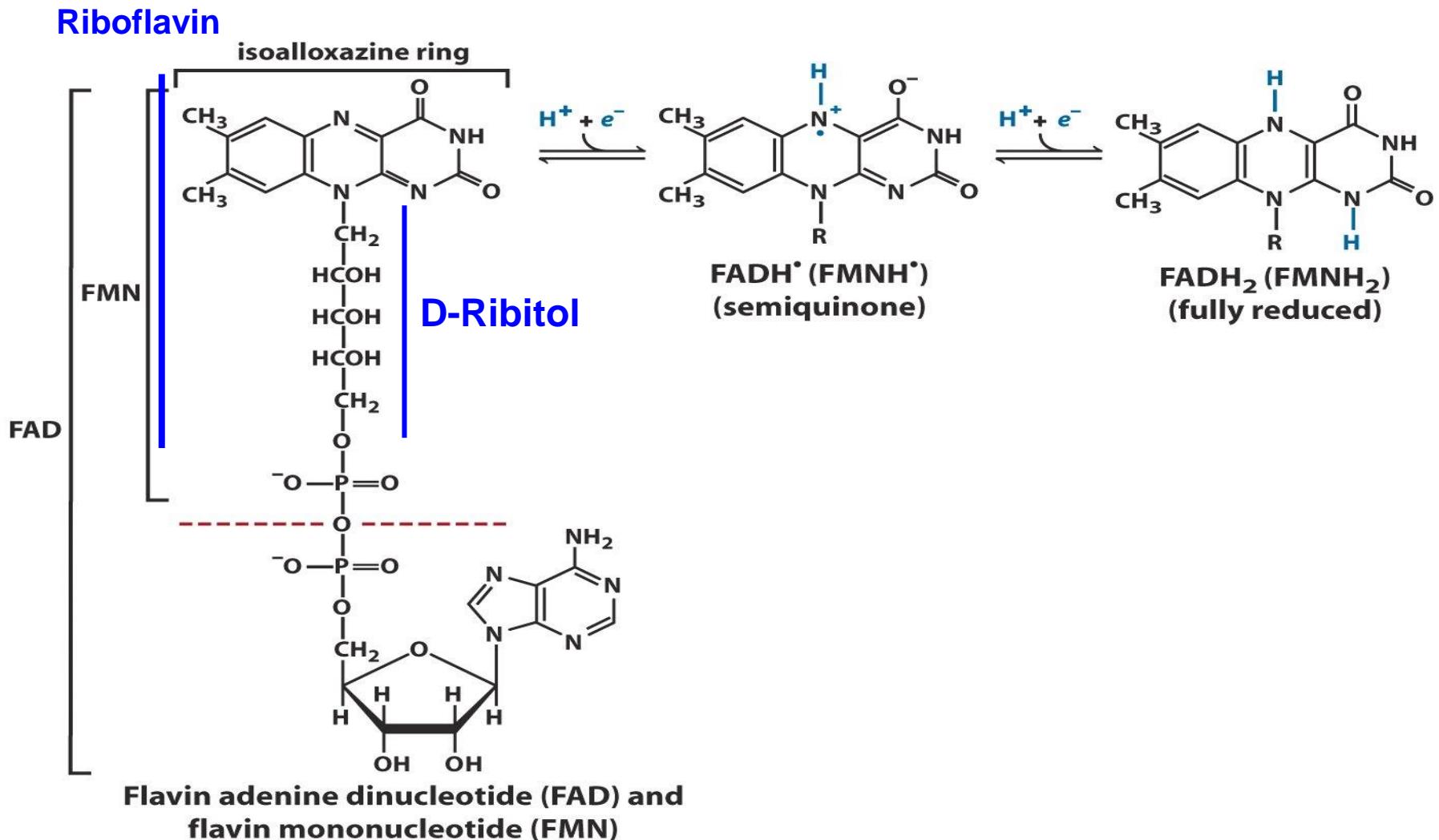


TABLE 13–9 Some Enzymes (Flavoproteins)
That Employ Flavin Nucleotide Coenzymes

| <i>Enzyme</i> | <i>Flavin nucleotide</i> | <i>Text page(s)</i> |
|------------------------------------|--------------------------|---------------------|
| Acyl-CoA dehydrogenase | FAD | 638 |
| Dihydrolipoyl dehydrogenase | FAD | 605 |
| Succinate dehydrogenase | FAD | 612 |
| Glycerol 3-phosphate dehydrogenase | FAD | 714–715 |
| Thioredoxin reductase | FAD | 869 |
| NADH dehydrogenase (Complex I) | FMN | 696–697 |
| Glycolate oxidase | FMN | 767 |